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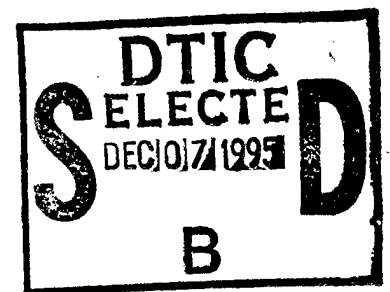
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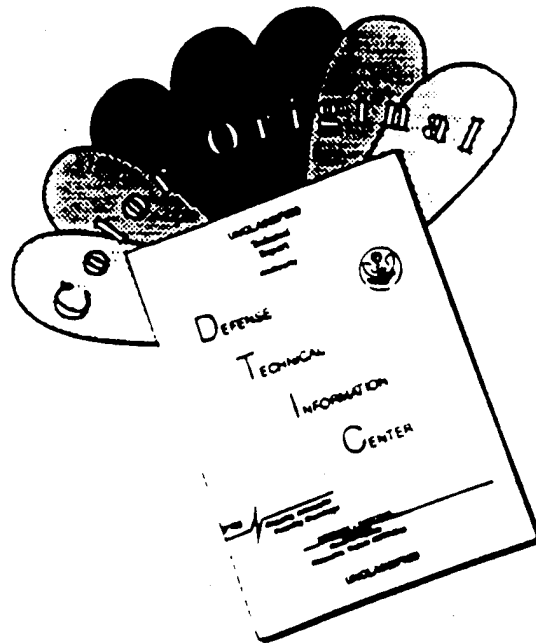
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Gene Therapy of Breast Cancer: Studies of Selection
Promoter/Enhancer-Modified Vectors to Deliver Suicide
Genes

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The high molecular weight mucin-like glycoprotein, DF3 (MUC1), is overexpressed in the majority of human breast cancers. Our ongoing studies demonstrate that replication of defective recombinant adenoviral vectors, containing the DF3 promoter (bps -725 to +31), can be used to express β -galactosidase (Ad.DF3- β gal) and the herpes simplex virus thymidine kinase (HSV-tk) gene (Ad.DF3-tk) in DF3 positive breast carcinoma cell lines. In vivo experiments using breast tumor implants in nude mice injected with Ad.DF3- β gal demonstrated that expression of the β -galactosidase gene is limited to DF3-positive breast cancer xenografts. Moreover, in an intraperitoneal breast cancer metastases model, we show that i.p. injection of Ad.DF3-tk followed by GCV treatment results in inhibition of tumor growth. These results demonstrate that utilization of the DF3 promoter in an adenoviral vector can confer selective expression of heterologous genes in breast cancer cells in vitro and in vivo.

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**Gene Therapy of Breast Cancer: Studies of selection
Promoter/Enhancer-modified Vectors to Deliver
Suicide Genes**

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INTRODUCTION

Most patients with metastatic breast cancer succumb to their disease. Chemotherapy is often standard treatment for these patients; however, effectiveness of the available chemotherapeutic drugs is limited by lack of selectivity and a narrow therapeutic window. Gene therapy is a potentially novel approach to cancer treatment. In this context, transfer of suitable genetic material into a specific cell type (either tumor or host) can be used to alter the phenotype of the target cell. One such strategy is based on direct transfer of a "suicide gene" which encodes an enzyme such as herpes simplex thymidine kinase (HSV-tk) that can activate a prodrug within tumor cells and thereby render the tumor cells sensitive to agents which are otherwise nontoxic to the cell. For example, ganciclovir (GCV) is a nucleoside that is nontoxic to mammalian cells, but is lethal following phosphorylation in cells that express HSV-tk (1-4). While gene therapy may provide a new therapeutic approach to breast cancer treatment, clinical efficacy will probably require a gene delivery system which possesses both high gene transduction efficiency and target cell specificity.

Human adenoviruses are non-enveloped double-stranded DNA viruses with a genomic size of approximately 36 kb (5,6). The E1 gene deleted adenoviruses are replication defective and can be grown in a packaging cell line transformed with the E1a and E1b genes (7). Adenoviral vectors deleted at the E1 and E3 regions are capable of accommodating DNA inserts up to 8 kb (8,9). Moreover, adenovirus-mediated gene transfer is a highly efficient

means for delivery of genetic material into a wide spectrum of cells both in vitro and in animals. Although recombinant adenoviruses hold promise for in vivo gene therapy and are being tested clinically (10), one of the limitations of this vector system for cancer therapy may be the non-specific transduction of therapeutic genes into non-target cells. One strategy to circumvent this limitation would be to use a tumor-tissue specific/selective promoter or enhancer to direct the expression of a therapeutic gene in the desired target cells.

DF3 antigen (also designated MUC1 and episialin) is a member of a family of high molecular weight glycoproteins which are aberrantly overexpressed in most human breast cancers. We have previously shown that monoclonal antibody MAb DF3, prepared against a membrane-enriched extract of a human breast carcinoma metastatic to liver, reacts with over 75% of primary human breast carcinomas (11). Other studies have shown that overexpression of the DF3 gene in human MCF-7 (12) and ZR-75 breast cancer cells (13) is regulated at the transcriptional level. Recent cloning and characterization of the 5' flanking region of DF3 gene has demonstrated that the DF3 gene expression is mainly regulated by sequences between positions -598 and -485 base pairs upstream to the transcription start site (14). This region contains a potential AP-2 site at position -587 (CCCCAGCC), an AP-3 site at position -596 (GGAAAGTCC), and SP1 sites at positions -582 (GGGCGG), -532 (GAGCGG), and -492 (GGGAGG). In addition, an unidentified cis element at position -505 to -485

(GGGAAGTGGTGGGGGGAGGGA) was found to specifically interact with nuclear proteins from MCF-7 breast cancer cells (14).

In our ongoing work, we have constructed replication defective adenoviral vectors containing the E. coli β -galactosidase gene (Ad.DF3- β -gal) or the HSV-tk gene (Ad.DF3-tk) under control of the DF3 promoter. We have evaluated expression of the reporter gene in DF3-positive and -negative cells in vitro and in vivo. We have also assessed the ability of Ad.DF3-tk to confer sensitivity to GCV in human breast cancer models in athymic nude mice. Our results suggest that an adenoviral vector system containing the DF3 promoter is capable of directing efficient and selective expression of heterologous genes in DF3-positive breast carcinomas.

BODY

Experimental Methods

Cell culture. The MCF-7, ZR-75-1, BT-20, and MDA-MB231 breast cancer cell lines were obtained from American Type Culture Collection (ATCC, Rockville, MD). The Hs578Bst myoepithelial cell line derived from normal breast tissue adjacent to a infiltrating ductal carcinoma (15) and the human T98G glioblastoma cell line were also obtained from ATCC. Cells were grown as monolayers in recommended culture medium supplemented with 10% heat-inactivated fetal bovine serum, 2 mM L-glutamine, 100 units/ml penicillin, and 100 µg/ml streptomycin. Cells were maintained in a 5% CO₂ humidified atmosphere.

Recombinant adenoviruses. Recombinant adenoviruses Ad.DF3-βgal and Ad.CMV-βgal derived from type 5 adenovirus (Ad 5), were produced by homologous recombination in the human embryonic kidney cell line 293 (7). The DF3 5' flanking region (-725 to -31) was inserted into Xho I and Spe I digested plasmid pCMVβgal (provided by Dr. R. Crystal, Cornell Medical Center, NY). The resulting plasmid pDF3βgal contains the E.coli β-galactosidase (lacZ) gene with the SV40 polyadenylation signal under the control of DF3 promoter and SV40 splice donor/acceptor signal, flanked by Ad 5 map units 0.0-1.3 and 9.3-17.3 (Fig. 1). To construct AD.DF3-tk, a 2.0 kb cDNA of HSV-tk (16) was used to replace the lacZ gene in the shuttle plasmid pDF3βgal (Fig. 1). The shuttle plasmids were cotransfected by calcium phosphate precipitation into 293 cells together with pJM17 plasmid containing the adenoviral genome (kindly provided by Dr. Graham, McMaster University, Ontario,

Canada). Recombinant adenovirus was isolated from a single plaque and expanded in 293 cells. The viral DNA was purified and analyzed by restriction enzyme digestion and by polymerase chain reaction (PCR). A pair of primers, 5'-TCTAGACTAGTGGACCCTAGGGTTCATCGGAG-3' and 5'-AACTCGAGGATTCAGGCAGGCGCTGGCT-3' was used to amplify and verify the presence of the DF3 promoter (-725 to -31) in the viral genome. Ad.CMV- β gal and AD.CMV-tk are structurally similar replication-deficient recombinant adenovirus in which the lacZ and HSV-tk genes, respectively, are under the control of cytomegalovirus (CMV) immediate-early promoter and enhancer. Large scale production of recombinant adenovirus was accomplished by growth in 293 cells and purification by double cesium gradient ultracentrifugation as described (17). The titers of purified adenovirus were determined by a spectrophotometer and by plaque assays.

Adenovirus infection in vitro. Twenty-four h after plating, cells were infected with adenovirus at a multiplicity of infection (MOI) of 10-50. 48 h later or at a specified time post infection, cells were evaluated for the expression of the reporter gene or evaluated for sensitivity to GCV.

Assays for β -galactosidase. Histochemical staining with 5-bromo-4-chloro-3-indolyl β -D-galactoside (X-Gal). Sections of fresh frozen tissue (12 μ m) or cells were fixed with 0.5% glutaraldehyde in phosphate-buffered saline (PBS) containing 1 mM $MgCl_2$ for 10 min, rinsed with PBS, and then incubated with X-Gal (1 mg/ml), 5 mM $K_3Fe(CN)_6$, 5 mM $K_4Fe(CN)_6$, 1 mM $MgCl_2$ in PBS for 4 h.

Fluorescence-activated cell sorting (FACS) analysis. (i)

Indirect immunofluorescent analysis of DF3 antigen. Cultured cells ($1-2 \times 10^6$) were washed extensively with 1% bovine serum albumin (BSA) in PBS and incubated with MAb DF3 (1 μ g/ml) or isotype identical control antibody mouse IgG (F-8765, Sigma Chemical Co., St. Louis, MO) at 4°C for 1 h, and then washed with 1% BSA/PBS. Cells were incubated with fluorescein isothiocyanate (FITC)-conjugated goat anti-mouse IgG (F2012, Sigma) at 4°C for 1 h, washed and analyzed on a Becton Dickinson FACScan. Intensity of fluorescence was determined for 10,000 cells and compared with the fluorescence obtained using a non-reactive immunoglobulin of the same isotype. (ii) FACS-GAL assay (18). Briefly, 1×10^6 cells were suspended in 50 μ l of serum-free culture medium and warmed to 37°C. An equal volume of 2 mM fluorescein di- β -D-galactopyranoside (FDG; Molecular Probes, Eugene, OR) was added to each aliquot of cells. The cells and FDG were mixed rapidly and incubated for 1 min at 37°C. Thereafter, cells were washed once with 4 ml ice-cold PBS and maintained in ice-cold PBS until analysis on a Becton Dickinson FACScan.

Assays for GCV sensitivity in vitro. Adenovirus (Ad)-infected and noninfected cells were plated at 4×10^4 cells/well in 6 well plates. GCV was added to the culture medium at various final concentrations (0-250 μ M). After 6-7 days of incubation, cells were washed with PBS and trypsinized. The number of viable cells were determined by trypan blue exclusion. Cell number was also assessed using a colorimetric cell proliferation (XTT) assay that measures the mitochondrial dehydrogenase activity of viable

cells . Results are expressed as a growth ratio of the number of cells in plates containing drugs as a percentage of that in the corresponding drug-free controls.

In vivo gene transfer to human breast cancer xenografts.

Female athymic nude mice (Swiss-nu/nu, Taconic, Germantown, NY), 20-25 g, were used. For mice bearing MCF-7 or ZR-75-1 tumors, a single pellet of 17, β -estradiol (1.7 mg/60-day release, Innovative Research, Toledo, OH) was implanted subcutaneously one day before tumor inoculation. i) Subcutaneous tumor model. Cells (MCF-7, ZR-75-1 and MDA-MB231) in exponential growth phase (1×10^7 in 0.2 ml) were injected subcutaneously in the flanks of the animals. At 4 to 6 weeks after tumor implantation, up to 5×10^8 plaque-forming units (pfu) of purified recombinant adenovirus in 20 μ l were injected into MCF-7, ZR-75-1, and MDA-MB231 xenografts, and into limb skeletal muscle. A Hamilton syringe with a 26 gauge needle was used for injection. The needle was coated with fine charcoal particles to mark the needle tract in order to verify co-localization of the expression of the reporter gene with the viral injection. Three days after adenoviral infection, the animals were sacrificed and expression of the reporter gene in tumor xenografts and host tissues was evaluated. ii) Intraperitoneal tumor model. MCF-7 cells (5×10^6) were injected i.p. for the development of intraperitoneal tumors (day 0). On days 4 and 5 after tumor cell injection, 1×10^9 pfu adenovirus in 0.5 ml PBS were injected i.p. into the mice. On day 7, the animals were treated with either saline or GCV (125 mg/kg) daily for 4 days by i.p. injection. The animals were sacrificed at 5-6 weeks after

tumor inoculation. Tumors were collected and weighed for each animal.

Results

Selective expression of β -galactosidase in human cell lines in vitro. A replication defective recombinant adenovirus was constructed that contains the β -galactosidase gene under control of the DF3 promoter (Ad.DF3- β gal; Fig. 1). Expression of β -galactosidase was evaluated in Ad.DF3- β gal-infected MCF-7, ZR-75-1, BT-20, and MDA-MB231 breast cancer cells. In addition, Hs578Bst, a myoepithelial cell line derived from normal breast tissue and T98G, a human glioblastoma cell line were used in these studies. Each cell line was infected with either Ad.DF3- β gal or Ad.CMV- β gal at a MOI of 50. Expression of β -galactosidase was observed in MCF-7, ZR-75-1, and BT-20 cells infected with Ad.DF3- β gal (Figs. 2A, B, C), whereas little if any β -galactosidase activity could be detected in similarly infected Hs578Bst, MDA-MB231 and T98G cells (Figs. 2D, E, F). In contrast, all of these cell lines showed strong expression of β -galactosidase when infected with Ad.CMV- β gal (Figs. 2G, H, I) in which the reporter gene is under control of the CMV early promoter and enhancer. MCF-7 cells infected with Ad.DF3- β gal at an MOI of 50 exhibited highest β -galactosidase activity at day 3 to day 7 post infection. Transgene expression gradually decreased to approximately 15% of maximum at 2 weeks post infection (data not shown).

Expression of β -galactosidase in Ad.DF3- β gal infected cells correlates with the expression of DF3. To assess whether there is a correlation between DF3 expression and capability of these cells to express β -galactosidase after Ad.DF3- β gal infection, we examined the presence of DF3 antigen in MCF-7, ZR-75-1, and Hs578Bst cells

by FACS analysis. MCF-7 and ZR-75-1 cells exhibited strong reactivity with MAb DF3, a monoclonal antibody against DF3 antigen, while little if any MAb DF3 binding was detectable with Hs578Bst cells (Fig. 3A). Using the FACS-GAL assay, all MCF-7 and ZR-75-1 cells infected with Ad.DF3- β gal appear to express β -galactosidase with an increase in mean fluorescent intensity of up to 35-fold (Fig. 3B). In contrast, there was little if any expression of β -galactosidase in Ad.DF3- β gal-infected Hs578Bst cells (Fig. 3B). These findings support a correlation between the presence of cellular DF3 antigen and expression of β -galactosidase in Ad.DF3- β gal-infected cells.

Ad.DF3-tk sensitizes DF3-positive MCF-7 and ZR-75-1 breast cancer cells to GCV in vitro. Given the finding that the DF3 promoter can direct selective expression of a reporter gene, we replaced the β -galactosidase gene in Ad.DF3- β gal with HSV-tk. To determine whether Ad.DF3-tk can confer sensitivity to GCV, MCF-7 and ZR-75-1 cells were transduced with Ad.DF3-tk at MOIs of 10 and 50. Ad.CMV-tk was used in order to assess HSV-tk gene expression under control of the different promoters. Infection with Ad.DF3-tk had little effect on cell viability (Fig. 4). Moreover, Ad.DF3-tk transduction conferred sensitivity of both MCF-7 and ZR-75 cells to GCV (Figs. 4A, 4B), while nontransduced cells or cells transduced with Ad.CMV- β gal or Ad.DF3- β gal (data not shown) were insensitive to GCV (Fig. 4). The degree of Ad.DF3-tk-mediated GCV sensitivity was comparable to that obtained with Ad.CMV-tk. In contrast, when Ad.DF3-tk and Ad.CMV-tk were used to infect DF3-

negative Hs578Bst epithelial cells, only Ad.CMV-tk infected cells were sensitive to GCV (Fig. 4C).

In vivo targeted gene expression in human breast cancer xenografts. To ascertain whether Ad.DF3- β gal can confer selective expression of β -galactosidase in vivo, we injected up to 5×10^8 pfu of Ad.DF3- β gal into MCF-7 and ZR-75-1 cells grown as subcutaneous xenografts in athymic nude mice. Three days after adenovirus infection, the tumors were excised and assayed for β -galactosidase activity. Expression of β -galactosidase was distributed extensively along the needle tract of Ad.DF3- β gal injection (as indicated by the co-localized charcoal particles) in both MCF-7 and ZR-75-1 tumor nodules (Fig. 5A and B). Blue staining was detectable within the tumor mass, but not in the surrounding normal tissue (Fig. 5B). In contrast, when Ad.DF3- β gal was injected into DF3-negative MDA-MB231 tumors grown in nude mice, there was no detectable β -galactosidase staining along the needle tract (Fig. 5C). Intramuscular injections of Ad.DF3- β gal also resulted in no detectable β -galactosidase activity (Fig. 5D), while similar injections of Ad.CMV- β gal into skeletal muscle was associated with strong expression of β -galactosidase (Fig. 5E).

β -galactosidase expression after systemic administration of Ad.DF3- β gal and Ad.CMV- β gal. To further evaluate adenovirus-mediated gene transfer to different tissues in vivo, 2×10^9 pfu of either Ad.CMV- β gal or Ad.DF3- β gal was injected via tail vein. Mice were sacrificed 4 days after the injection and sections of the liver, spleen, and lung were stained for β -galactosidase activity. Systemic injection of Ad.CMV- β gal resulted in expression of β -

galactosidase in the liver parenchyma, in the splenic red pulp, and diffusely in the lung (Fig. 6A). By contrast, there was no detectable β -galactosidase staining in these tissues in mice injected with Ad.DF3- β gal. Several foci of β -galactosidase staining were detected in MCF-7 tumors of animals that received systemic Ad.DF3- β gal (Fig. 6B). However, i.p. injection of Ad.DF3- β gal into mice bearing intraperitoneal MCF-7 tumor was associated with extensive expression of β -galactosidase in the tumor (Fig. 6C).

Treatment of intraperitoneal breast tumor. In order to evaluate the therapeutic efficacy of Ad.DF3-tk in human DF3-positive breast cancer cells in vivo, nude mice were inoculated i.p. with MCF-7 cells. These mice developed tumor masses throughout the peritoneal cavity and 2-4 ml of bloody ascites that contained tumor cells (Fig. 7A). MCF-7 tumor bearing mice were treated with Ad.DF3- β gal + saline, Ad.DF3- β gal + GCV, Ad.DF3-tk + saline and Ad.DF3-tk + GCV. Adenoviruses were injected i.p. on day 4 after tumor inoculation. GCV or saline injections were then administered on day 7 after tumor inoculation. Untreated mice and those treated with Ad.DF3- β gal or Ad.DF3-tk + saline developed multiple intraperitoneal tumors with bloody ascites. In contrast, there was no grossly identifiable tumor mass or only a few small tumor clusters with no apparent ascites in the Ad.DF3-tk/GCV-treated mice (Figs. 7B and 7C).

Discussion

The results presented here demonstrate that the DF3 gene promoter in the context of a recombinant adenoviral vector can confer selective expression of a reporter gene in DF3-positive human breast cancer cells in vitro and in vivo. Moreover, we demonstrate that adenovirus-mediated transduction of HSV-tk under control of the DF3 promoter can confer selective sensitivity of DF3 positive cells to GCV. These findings support the use of tumor-selective promoters to target expression of therapeutic genes in adenovirus-mediated gene therapy.

Currently available in vivo gene delivery systems generally lack target cell specificity. Non-viral approaches, such as ligand-DNA complexes mediated gene transduction through receptor endocytosis have been reported to deliver genes through asialoglycoprotein receptors (19,20). This approach, however, is largely hindered by the relatively low gene transduction efficiency due to the endosomal/lysosomal trapping of the DNA complex. Although other approaches such as the use of DNA liposome complexes (21,22) and direct injection of DNA (23) have been applied for direct in vivo gene transfer, they are also limited by relative low gene transduction or restricted target tissues. Retroviral vectors have been used extensively for gene transfer, particularly for ex vivo gene therapy (24-27). However, application of retroviruses for in vivo gene therapy may be limited by low viral titers and the dependence on target cell replication. In addition, random chromosomal integration of retroviruses may raise some potential safety concerns.

Recently, recombinant adenoviruses have been employed as a highly efficient approach for in vitro and in vivo gene transfer. Adenovirus-mediated gene transduction is independent of cell replication and the virus is capable of infecting a broad spectrum of eukaryotic cells. However, one limitation of this vector system for clinical application may be the non-specific transduction of therapeutic genes into cells other than the target cells. This limitation may be especially critical in the case of gene therapy for cancer, in which "suicide" genes are to be delivered into the tumor cells. Rat brain gliomas have been treated by intracranial injection of adenovirus carrying HSV-tk driven by the constitutive RSV promoter (28). Since gliomas are not metastatic and reside within a mitotically quiescent central nervous system, the transduction of a prodrug-activating gene, like HSV-tk, by local injection of adenovirus, is sufficient to achieve tumor response. However, in systemic tumors, successful therapy may require both efficiency and specificity of gene expression in target cells. Therefore, one potential strategy is to design an adenoviral vector in which a therapeutic gene can be expressed selectively in tumor cells with little or no expression in normal cells. Although adenovirus infects a wide range of cells, the present studies demonstrate that expression of a heterologous gene under the control of DF3 promoter in the context of an adenovirus vector is limited to DF3-positive breast cancer cells.

To date, only a few tumor tissue-specific or -selective DNA regulatory sequences have been identified. Moreover, most studies

of selective promoters to target tumor cells were performed in cell culture or ex vivo models. Selective expression in melanoma cells has been reported using plasmids containing the promoters of the genes encoding tyrosinase and tyrosinase related proteins (29,30). The promoter of the α -fetoprotein gene (31), the promoter of the oncogene ERBB2 (32), and recently the enhancer sequence (-598 to -485) of the DF3 gene (33) have been explored in the context of retroviral vectors to direct expression of prodrug activating enzymes and to confer selective tumor killing in a number of cancer cell lines in vitro. However, to our knowledge, there are no previous reports of using a tumor-selective promoter in the context of an adenovirus vector to selectively kill tumor cells in vivo. In the present studies, we have demonstrated an efficient and selective approach to target expression of heterogenous genes in DF3-positive breast cancer cells.

CONCLUSIONS

The results suggest that recombinant adenoviruses containing the DF3 promoter and therapeutic genes could have applicability in gene therapy of breast cancer. Currently, recombinant adenoviral vectors containing the DF3 promoter and other candidate therapeutic genes including cytochrome P450 (34,35), and cytosine deaminase (36,37) are being developed. Of importance, E1-deleted adenoviral vectors used in our studies are referred to as "first generation" vectors which have been associated with inherent cytotoxicity and immunogenicity. The engineering of newer versions of vector by further crippling the viral genome (such as E2a and E4) and by enhancing expression of the transgenes may eventually lead to a vector system that is practical for human gene therapy.

REFERENCES

1. Moolten, F. L., and J. M. Wells. 1990. Curability of tumors bearing herpes thymidine kinase genes transferred by retroviral vectors. *J. Natl. Cancer Inst.* 82:297-300.
2. Ezzeddine, Z. D., R. L. Martuza, D. Platika, M. P. Short, A. Malick, B. Choi, and X. O. Breakefield. 1991. Selective killing of glioma cells in culture and in vivo by retrovirus transfer of the herpes simplex virus thymidine kinase gene. *New Biol.* 3:608-14.
3. Culver, K. W., Z. Ram, S. Wallbridge, H. Ishii, E. H. Oldfield, and R. M. Blaese. 1992. In vivo gene transfer with retroviral vector-producer cells for treatment of experimental brain tumors. *Science* 256:1550-2.
4. Ram, Z., K. W. Culver, S. Walbridge, R. M. Blaese, and E. H. Oldfield. 1993. In situ retroviral-mediated gene transfer for the treatment of brain tumors in rats. *Cancer Res.* 53:83-8.
5. Ginsberg, H. S. 1984. The Adenoviruses. Plenum Press, New York.
6. Horwitz, M. S. 1990. Adenoviridae. In *Virology*. B. N. Fields and D. M. Knipe, editors. Raven Press, New York.
7. Graham, F. L., J. Smiley, W. C. Russel, and R. Nairn. 1977. Characteristics of a human cell line transformed by DNA from human adenovirus type 5. *J. Gen. Virol.* 36:59-72.
8. Haj-Ahmad, Y., and F. L. Graham. 1986. Characterization of an adenovirus type 5 mutant carrying embedded inverted terminal repeats. *Virology* 153:22-34.

9. Bett, A. J., W. Haddara, L. Prevec, and F. L. Graham. 1994. An efficient and flexible system for construction of adenovirus vector with insertions or deletions in early regions 1 and 3. *Proc. Natl. Acad. Sci. USA* 91:8802-8806.
10. Crystal, R. G., N. G. McElvaney, M. A. Rosenfeld, C. Chu, A. Mastrangeli, J. G. Hay, S. T. Brody, H. A. Jaffe, N. T. Eissa, and C. Danel. 1994. Administration of an adenovirus containing the human CFTR cDNA to the respiratory tract of individuals with cystic fibrosis. *Nature Genet.* 8:42-50.
11. Kufe, D., G. Inghirami, M. Abe, D. Hayes, H. Justi-Wheeler, and J. Schlom. 1984. Differential reactivity of a novel monoclonal antibody (DF3) with human malignant versus benign breast tumors. *Hybridoma* 3:223-32.
12. Abe, M., and D. Kufe. 1990. Transcriptional regulation of DF3 gene expression in human MCF-7 breast carcinoma cells. *J. Cell. Physiol.* 143:226-31.
13. Kovarik, A., N. Peat, D. Wilson, S. J. Gendler, and J. Taylor-Papadimitriou. 1993. Analysis of the tissue-specific promoter of the MUC1 gene. *J. Biol. Chem.* 268:9917-26.
14. Abe, M., and D. Kufe. 1993. Characterization of cis-acting elements regulating transcription of the human DF3 breast carcinoma-associated antigen (MUC1) gene. *Proc. Natl. Acad. Sci. USA* 90:282-6.
15. Hackett, A. J., H. S. Smith, E. L. Springer, R. B. Owens, W. A. Nelson-Rees, J. L. Riggs, and M. B. Gardner. 1977. Two syngeneic cell lines from human breast tissue: the aneuploid

mammary epithelial (Hs578T) and the diploid myoepithelial (Hs578Bst) cell lines. *J. Natl. Cancer Inst.* 58:1795-1806.

16. Colbere-Garapin, F., S. Chousterman, F. Horodniceanu, P. Kourilsky, and A. C. Garapin. 1979. Cloning of the active thymidine kinase gene of herpes simplex virus type 1 in *Escherichia coli* K-12. *Proc. Natl. Acad. Sci. U.S.A.* 76:3755-3759.

17. Graham, F. L., and L. Prevec. 1991. Manipulation of adenovirus vectors. In *Methods in Molecular Biology: Gene transfer and expression protocols*. E. J. Murray, editor. The Human Press Inc., Clifton, NJ.

18. Nolan, G. P., S. Fiering, J. F. Nicolas, and L. A. Herzenberg. 1988. Fluorescence-activated cell analysis and sorting of viable mammalian cells based on beta-D-galactosidase activity after transduction of *Escherichia coli* lacZ. *Proc. Natl. Acad. Sci. USA* 85:2603-7.

19. Wu, G. Y., and C. H. Wu. 1988. Receptor-mediated gene delivery and expression in vivo. *J. Biol. Chem.* 263:14621-4.

20. Wu, G. Y., J. M. Wilson, F. Shalaby, M. Grossman, D. A. Shafritz, and C. H. Wu. 1991. Receptor-mediated gene delivery in vivo. Partial correction of genetic analbuminemia in Nagase rats. *J. Biol. Chem.* 266:14338-42.

21. Nabel, E. G., G. Plautz, and G. J. Nabel. 1990. Site-specific gene expression in vivo by direct gene transfer into the arterial wall. *Science* 249:1285-8.

22. Nabel, E. G., D. Gordon, Z. Y. Yang, L. Xu, H. San, G. E. Plautz, B. Y. Wu, X. Gao, L. Huang, and G. J. Nabel. 1992. Gene

transfer in vivo with DNA-liposome complexes: lack of autoimmunity and gonadal localization. *Hum. Gene Ther.* 3:649-56.

23. Wolff, J. A., R. W. Malone, P. Williams, W. Chong, G. Acsadi, A. Jani, and P. L. Felgner. 1990. Direct gene transfer into mouse muscle in vivo. *Science* 247:1465-8.

24. Culver, K., K. Cornetta, R. Morgan, S. Morecki, P. Aebersold, A. Kasid, M. Lotze, S. A. Rosenberg, W. F. Anderson, and R. M. Blaese. 1991. Lymphocytes as cellular vehicles for gene therapy in mouse and man. *Proc. Natl. Acad. Sci. USA* 88:3155-9.

25. Wilson, J. M., M. Grossman, S. E. Raper, J. Baker Jr., R. S. Newton, and J. G. Thoene. 1992. Ex vivo gene therapy of familial hypercholesterolemia. *Hum. Gene Ther.* 3:179-222.

26. Dranoff, G., E. Jaffee, A. Lazenby, P. Golumbek, H. Levitsky, K. Brose, V. Jackson, H. Hamada, D. Pardoll, and R. C. Mulligan. 1993. Vaccination with irradiated tumor cells engineered to secrete murine granulocyte-macrophage colony-stimulating factor stimulates potent, specific, and long-lasting anti-tumor immunity. *Proc. Natl. Acad. Sci. USA* 90:3539-43.

27. Jaffee, E. M., G. Dranoff, L. K. Cohen, K. M. Hauda, S. Clift, F. F. Marshall, R. C. Mulligan, and D. M. Pardoll. 1993. High efficiency gene transfer into primary human tumor explants without cell selection. *Cancer Res.* 53:2221-6.

28. Chen, S. H., H. D. Shine, J. C. Goodman, R. G. Grossman, and S. L. Woo. 1994. Gene therapy for brain tumors: regression of experimental gliomas by adenovirus-mediated gene transfer in vivo. *Proc. Natl. Acad. Sci. USA* 91:3054-7.

29. Vile, R. G., and I. R. Hart. 1993. In vitro and in vivo targeting of gene expression to melanoma cells. *Cancer Res.* 53:962-7.
30. Vile, R. G., and I. R. Hart. 1993. Use of tissue-specific expression of the herpes simplex virus thymidine kinase gene to inhibit growth of established murine melanomas following direct intratumoral injection of DNA. *Cancer Res.* 53:3860-4.
31. Huber, B. E., C. A. Richards, and T. A. Krenitsky. 1991. Retroviral-mediated gene therapy for the treatment of hepatocellular carcinoma: an innovative approach for cancer therapy. *Proc. Natl. Acad. Sci. USA* 88:8039-43.
32. Harris, J. D., A. A. Gutierrez, H. C. Hurst, K. Sikora, and N. R. Lemoine. 1994. Gene therapy for cancer using tumor-specific prodrug activation. *Gene Therapy* 1:170-175.
33. Manome, Y., M. Abe, M. F. Hagen, H. A. Fine, and D. W. Kufe. 1994. Enhancer sequences of the DF3 gene regulate expression of the herpes simplex virus thymidine kinase gene and confer sensitivity of human breast cancer cells to ganciclovir. *Cancer Res.* 54:5408-5413.
34. Wei, M. X., T. Tamiya, M. Chase, E. J. Boviatsis, T. K. H. Chang, N. W. Kowall, F. H. Hochberg, D. J. Waxman, X. O. Breakefield, and E. A. Chiocca. 1994. Experimental tumor therapy in mice using the cyclophosphamide-activating cytochrome P4502B1 gene. *Hum. Gene Ther.* 5:969-78.
35. Chen, L., and D. J. Waxman. 1995. Intratumoral activation and enhanced chemotherapeutic effect of oxazaphosphorines following cytochrome P450 gene transfer: development of a combined

chemotherapy/cancer gene therapy strategy. *Cancer Res.* 55:581-589.

36. Mullen, C. A., M. Kilstrup, and R. M. Blaese. 1992. Transfer of the bacterial gene for cytosine deaminase to mammalian cells confers lethal sensitivity to 5-fluorocytosine: a negative selection system. *Proc. Natl. Acad. Sci. USA* 89:33-7.

37. Huber, B. E., E. A. Austin, S. S. Good, V. C. Knick, S. Tibbels, and C. A. Richards. 1993. In vivo antitumor activity of 5-fluorocytosine on human colorectal carcinoma cells genetically modified to express cytosine deaminase. *Cancer Res.* 53:4619-26.

Appendix

Figure 1. Schematic representation of the recombinant Ad.DF3- β gal and Ad.DF3-tk adenoviral vectors. Ad.DF3- β gal was constructed by insertion of a β -galactosidase (lacZ) expression cassette containing the DF3 promoter and SV40 polyadenylation signal at the deleted E1 region Ad 5 (1.3-9.2 mu). Ad.DF3-tk is a structurally similar adenovirus in which the lacZ gene is replaced by the HSV-tk gene.

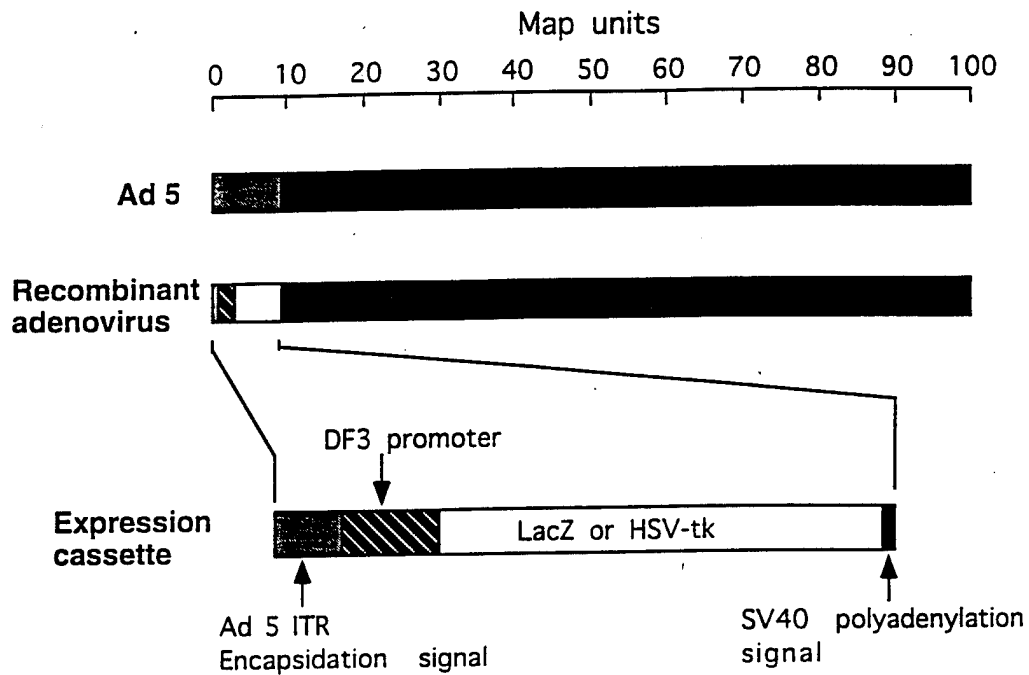


Figure 2. Expression of β -galactosidase in human cell lines infected with recombinant adenoviruses. Two days after adenovirus infection (MOI = 50), cells were fixed in 0.5% glutaraldehyde and stained with X-Gal for 4 h. MCF-7 (A), ZR-75-1 (B), BT-20 (C), Hs578Bst (D), MDA-MB231 (E), and T98G (F) cells infected with Ad.DF3- β gal. Hs578Bst (G), MDA-MB231 (H), and T98G (I) cells infected with Ad.CMV- β gal. All panels 100x magnification.

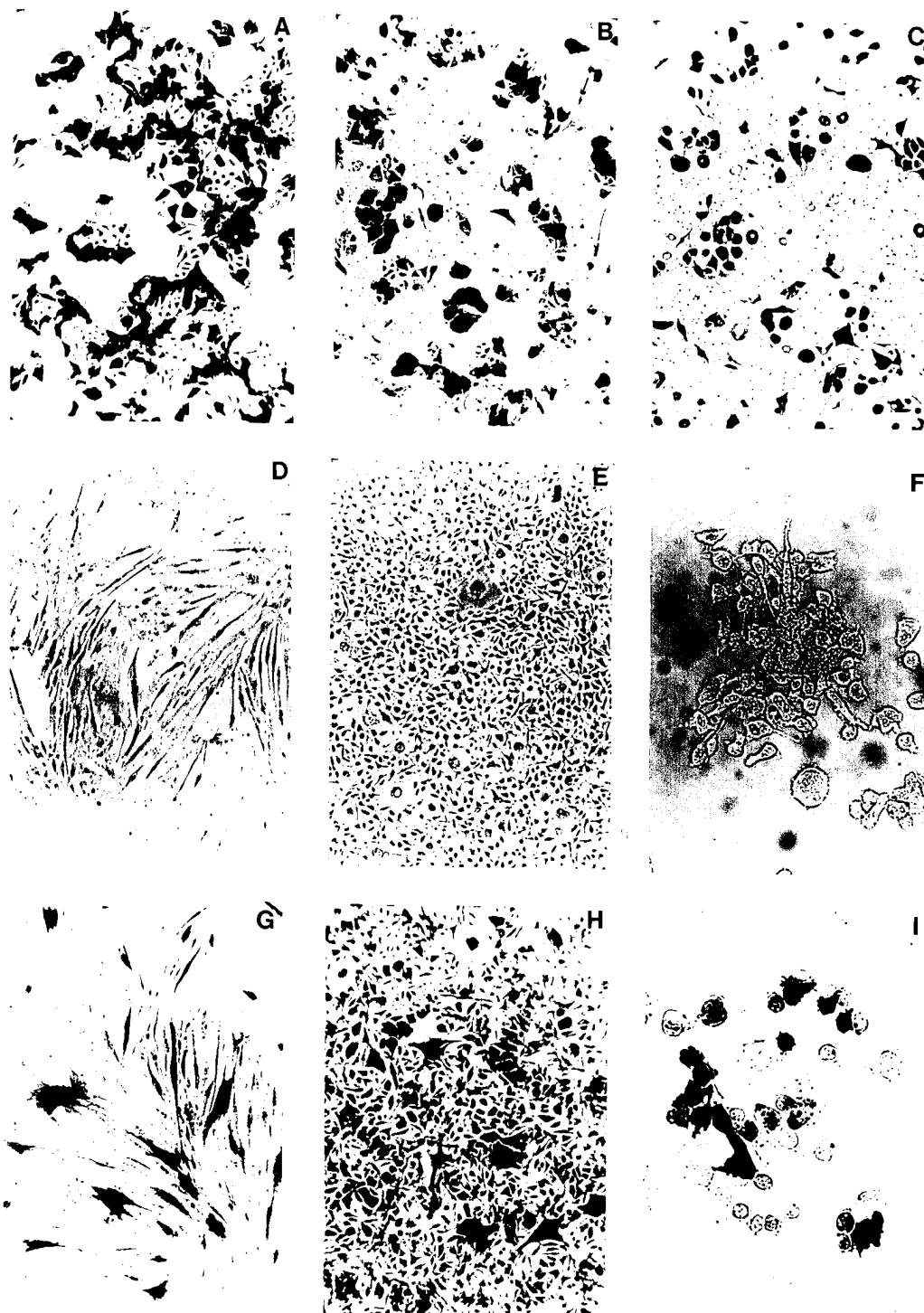


Figure 3. β -galactosidase expression in Ad.DF3- β gal infected cells correlates with expression of DF3 antigen. (A) FACS analysis to detect the presence of DF3 antigen in MCF-7, ZR-75-1 and Hs578Bst cell lines. Cells were incubated with mouse IgG as control (curve a) or Mab DF3 (curve b) and then FITC-conjugated anti-mouse IgG. (B) FACS-GAL assay to analyze β -galactosidase activity in uninfected cells (curve a) or Ad.DF3- β gal (MOI = 50) infected cells (curve b). Y-axis indicates relative number of cells, x-axis shows fluorescence intensity.

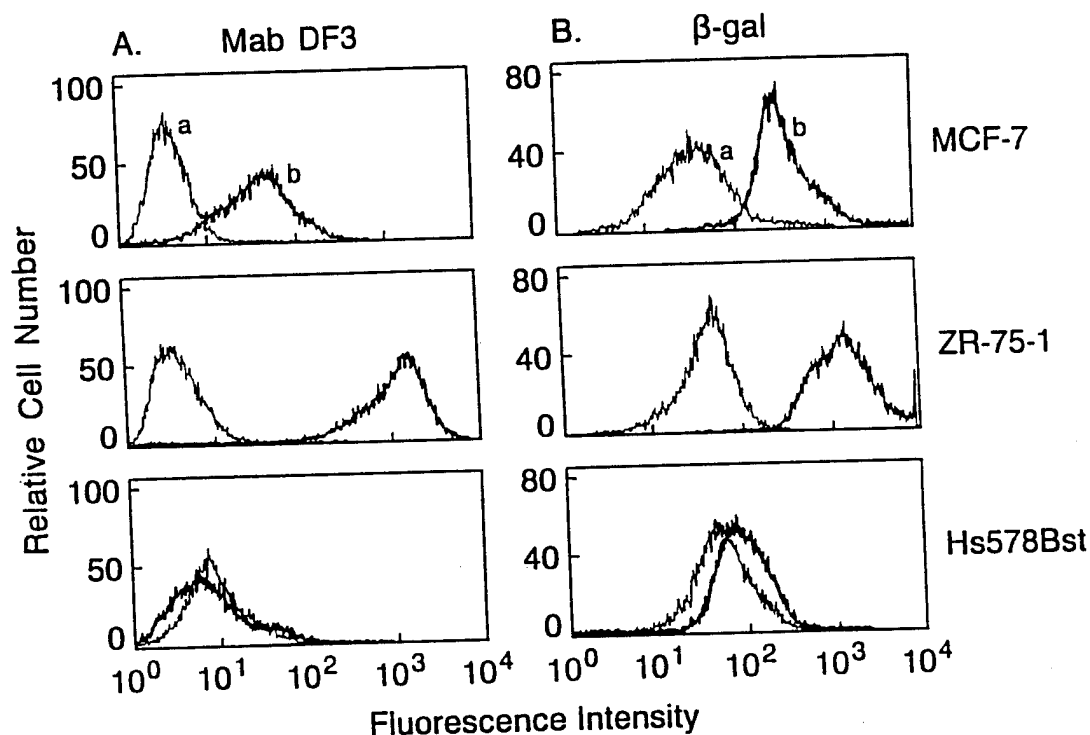


Figure 4. Ad.DF3-tk selectively sensitizes DF3-positive breast cancer cells to GCV. Cells were infected with Ad.DF3-tk or Ad.CMV-tk at MOIs of 10 to 50. Uninfected cells or cells infected with Ad.CMV- β gal were used as controls. 48 h later, cells were replated at 4×10^4 cells/well in 6-well plates. GCV was then added at concentrations ranging from 0 to 250 μ M. After 6-7 days of incubation, viable cells were counted using trypan blue exclusion. The effect of GCV on cell survival was expressed as the growth ratio, i.e. cell number in wells containing drug as a percentage of the corresponding drug free control (mean \pm SE for duplicate determinations). (—□—) no adenovirus; (—○—) Ad.CMV- β gal (MOI = 50); (—■—) Ad.DF3-tk (MOI = 10); (—▲—) Ad.DF3-tk (MOI = 50); (—●—) Ad.CMV-tk (MOI = 50). Similar results were obtained in three separate experiments.

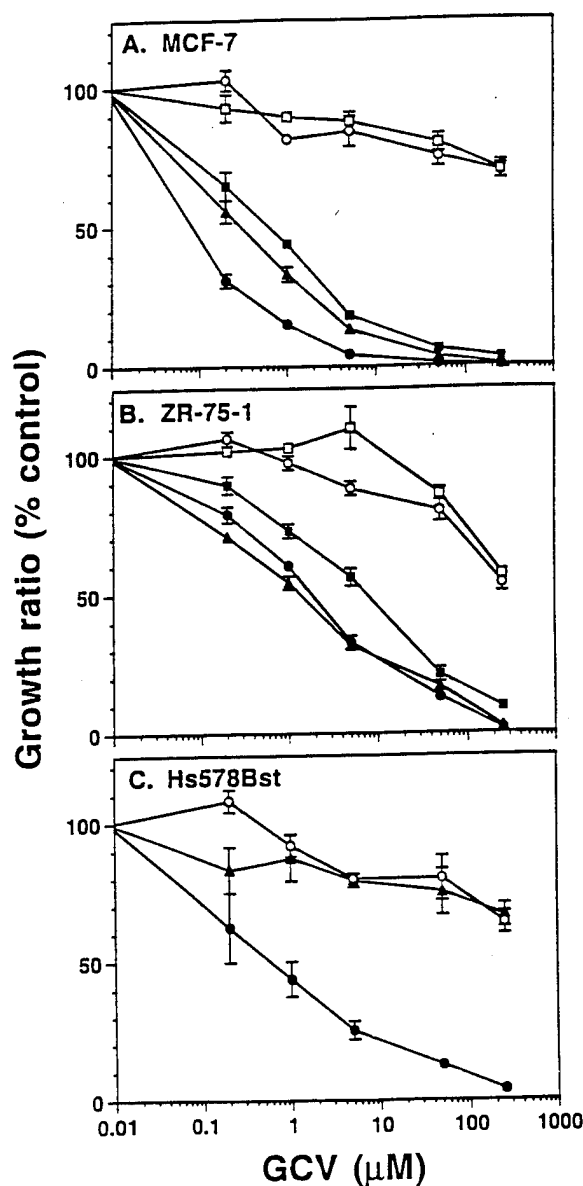


Figure 5. Ad.DF3- β gal mediated in vivo selective expression of β -galactosidase in breast cancer cells. MCF-7, ZR-75-1, and MDA-MB231 xenografts were grown subcutaneously in athymic nude mice. Adenovirus (5×10^8 pfu) was injected into the tumors or mouse limb skeletal muscle using a 26-gauge needle coated with charcoal particles (indicated by the arrowhead). The tissues were harvested 3 days after viral injection. The frozen sections (12 μ m) were fixed in 0.5% glutaraldehyde and stained for β -galactosidase activity with X-Gal for 4 h. MCF-7 (A), ZR-75-1 (B), and MDA-MB231 (C) injected with Ad.DF3- β gal. Mouse skeletal muscle injected with Ad.DF3- β gal (D) or with Ad.CMV- β gal (E). All panels 100x magnification.

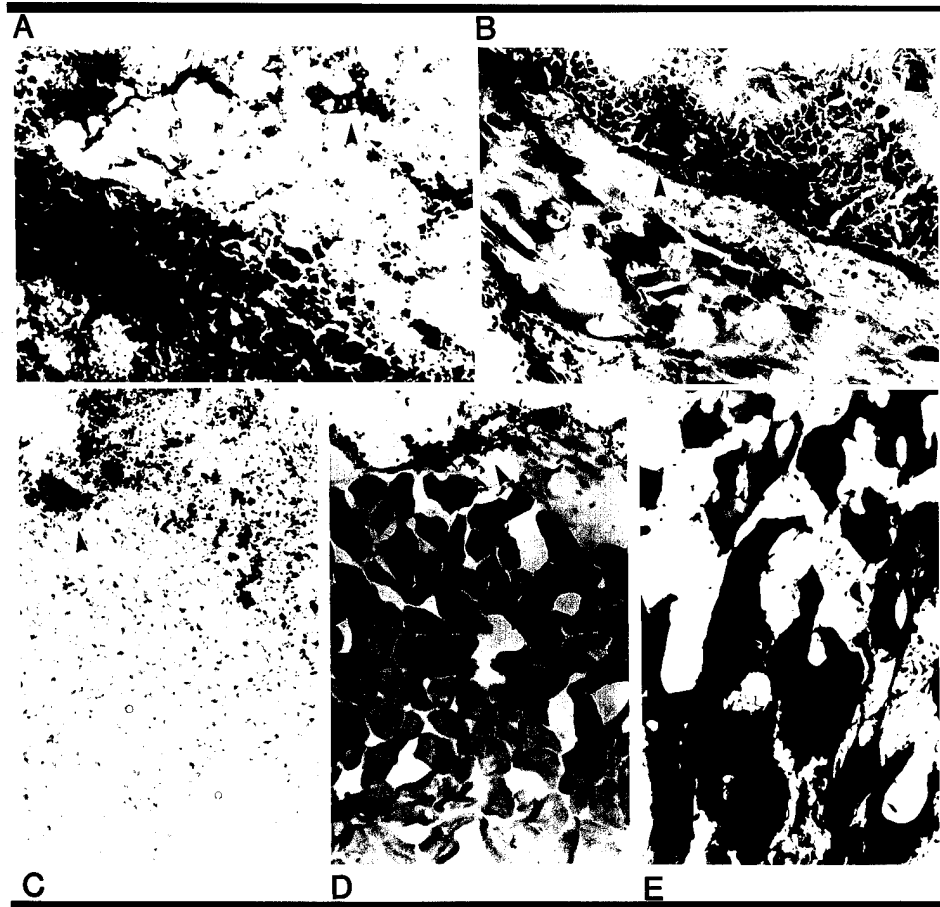


Figure 6. Distribution of β -galactosidase expression in mice injected systemically with recombinant adenoviruses. 4 days after adenovirus injection, sections (12 μ M) of the individual tissues were prepared and stained with X-Gal for β -galactosidase activity. (A) Mice were injected with 2×10^9 pfu of Ad.DF3- β gal or Ad.CMV- β gal in 0.1 ml PBS through tail vein. Expression of β -galactosidase was evaluated in liver, spleen, and lung. 40x magnification. (B) Mice bearing intraperitoneal MCF-7 tumors were given 2×10^9 pfu Ad.DF3- β gal intravenously. Tumor was evaluated for β -galactosidase expression. 100x magnification. (C) Mice bearing intraperitoneal MCF-7 tumors were given 2×10^9 pfu Ad.DF3- β gal i.p. Tumor was evaluated for β -galactosidase expression. 100x magnification.

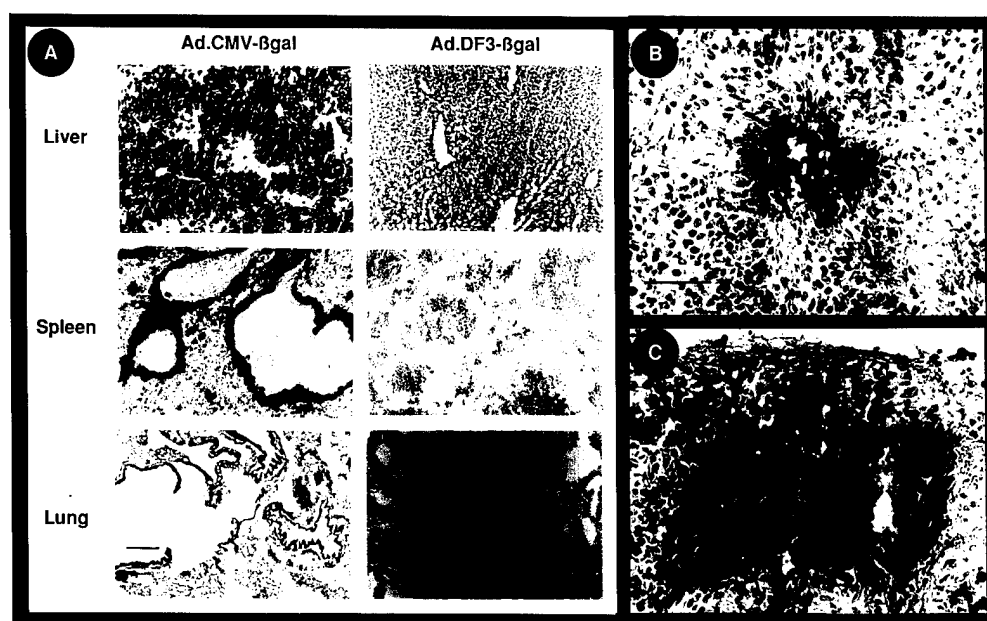


Figure 7. Treatment of intraperitoneal MCF-7 breast tumors. MCF-7 cells (5×10^6) were inoculated i.p. into nude mice. On days 4 and 5, mice received i.p. injections of Ad.DF3-tk or Ad.DF3- β gal (1×10^9 pfu) in 0.5 ml PBS. GCV treatment was initiated on day 7 at a dose of 125 mg/kg body weight daily via i.p. injection for 4 days. Six weeks after GCV treatment, animals were sacrificed for evaluation. (A) Intraperitoneal tumor in an untreated mouse, showing multiple solid tumors (arrows) and bloody ascites. (B) Mouse treated i.p. with Ad.DF3-tk/GCV showing no evident intraperitoneal tumor. (C) Tumor masses from each group were collected and weighed (mean \pm SE, $n = 6$). In the Ad.DF3-tk + GCV treatment group, two mice had no grossly visible tumor. Similar results were obtained in three separate experiments. Asterisk: significance ($p < 0.001$) compared to other groups.



C.
Tumor Weight (gram)

